

Resource Summary Report

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Anti-Vesicular Acetylcholine Transporter (VACHT) antibody produced in rabbit

RRID:AB_261875

Type: Antibody

Proper Citation

(Sigma-Aldrich Cat# V5387, RRID:AB_261875)

Antibody Information

URL: http://antibodyregistry.org/AB_261875

Proper Citation: (Sigma-Aldrich Cat# V5387, RRID:AB_261875)

Target Antigen: Vesicular Acetylcholine Transporter (VACHT) antibody produced in rabbit

Host Organism: rabbit

Clonality: polyclonal

Comments: Vendor recommendations: immunohistochemistry (formalin-fixed, paraffin-embedded sections): 1:1,000; Western Blot; Immunofluorescence; Immunohistochemistry

Antibody Name: Anti-Vesicular Acetylcholine Transporter (VACHT) antibody produced in rabbit

Description: This polyclonal targets Vesicular Acetylcholine Transporter (VACHT) antibody produced in rabbit

Target Organism: rat, human

Defining Citation: [PMID:22700183](#), [PMID:21031558](#), [PMID:17029265](#), [PMID:16871528](#)

Antibody ID: AB_261875

Vendor: Sigma-Aldrich

Catalog Number: V5387

Record Creation Time: 20241016T231229+0000

Record Last Update: 20241017T001428+0000

Ratings and Alerts

No rating or validation information has been found for Anti-Vesicular Acetylcholine Transporter (VAChT) antibody produced in rabbit.

No alerts have been found for Anti-Vesicular Acetylcholine Transporter (VAChT) antibody produced in rabbit.

Data and Source Information

Source: [Antibody Registry](#)

Usage and Citation Metrics

We found 11 mentions in open access literature.

Listed below are recent publications. The full list is available at [FDI Lab - SciCrunch.org](#).

Takeoka A, et al. (2019) Functional Local Proprioceptive Feedback Circuits Initiate and Maintain Locomotor Recovery after Spinal Cord Injury. Cell reports, 27(1), 71.

Valadão PAC, et al. (2018) Neuromuscular synapse degeneration without muscle function loss in the diaphragm of a murine model for Huntington's Disease. Neurochemistry international, 116, 30.

Wi?ckowska A, et al. (2018) Spinalization and locomotor training differentially affect muscarinic acetylcholine receptor type 2 abutting on ?-motoneurons innervating the ankle extensor and flexor muscles. Journal of neurochemistry, 147(3), 361.

Reyes C, et al. (2015) Distribution and innervation of putative peripheral arterial chemoreceptors in the red-eared slider (Trachemys scripta elegans). The Journal of comparative neurology, 523(9), 1399.

Suzuki DG, et al. (2015) A comparative examination of neural circuit and brain patterning between the lamprey and amphioxus reveals the evolutionary origin of the vertebrate visual center. The Journal of comparative neurology, 523(2), 251.

Reyes C, et al. (2014) Distribution and innervation of putative arterial chemoreceptors in the bullfrog (Rana catesbeiana). The Journal of comparative neurology, 522(16), 3754.

Spirovski D, et al. (2012) Brainstem galanin-synthesizing neurons are differentially activated by chemoreceptor stimuli and represent a subpopulation of respiratory neurons. The Journal

of comparative neurology, 520(1), 154.

Aizawa H, et al. (2012) Molecular characterization of the subnuclei in rat habenula. The Journal of comparative neurology, 520(18), 4051.

Bautista TG, et al. (2010) Cholinergic inputs to laryngeal motoneurons functionally identified in vivo in rat: a combined electrophysiological and microscopic study. The Journal of comparative neurology, 518(24), 4903.

Darrow KN, et al. (2006) Dopaminergic innervation of the mouse inner ear: evidence for a separate cytochemical group of cochlear efferent fibers. The Journal of comparative neurology, 498(3), 403.

Henny P, et al. (2006) Innervation of orexin/hypocretin neurons by GABAergic, glutamatergic or cholinergic basal forebrain terminals evidenced by immunostaining for presynaptic vesicular transporter and postsynaptic scaffolding proteins. The Journal of comparative neurology, 499(4), 645.