

Resource Summary Report

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Arginine-vasopressin specific neurophysin

RRID:AB_2313960

Type: Antibody

Proper Citation

(H. Gainer, National Institute of Neurological Disorders and Stroke Cat# PS41, RRID:AB_2313960)

Antibody Information

URL: http://antibodyregistry.org/AB_2313960

Proper Citation: (H. Gainer, National Institute of Neurological Disorders and Stroke Cat# PS41, RRID:AB_2313960)

Clonality: unknown

Antibody Name: Arginine-vasopressin specific neurophysin

Description: This unknown targets

Defining Citation: [PMID:20853513](https://pubmed.ncbi.nlm.nih.gov/20853513/)

Antibody ID: AB_2313960

Vendor: H. Gainer, National Institute of Neurological Disorders and Stroke

Catalog Number: PS41

Record Creation Time: 20231110T042049+0000

Record Last Update: 20241115T114414+0000

Ratings and Alerts

No rating or validation information has been found for Arginine-vasopressin specific neurophysin.

No alerts have been found for Arginine-vasopressin specific neurophysin.

Data and Source Information

Source: [Antibody Registry](#)

Usage and Citation Metrics

We found 12 mentions in open access literature.

Listed below are recent publications. The full list is available at [FDI Lab - SciCrunch.org](#).

Greenwood M, et al. (2024) Dimerization of hub protein DYNLL1 and bZIP transcription factor CREB3L1 enhances transcriptional activation of CREB3L1 target genes like arginine vasopressin. *Peptides*, 179, 171269.

Fernandez G, et al. (2023) Ghrelin Action in the PVH of Male Mice: Accessibility, Neuronal Targets, and CRH Neurons Activation. *Endocrinology*, 164(11).

Papazoglou I, et al. (2022) A distinct hypothalamus-to-? cell circuit modulates insulin secretion. *Cell metabolism*, 34(2), 285.

Basu S, et al. (2022) Secretagogen in the brain and pituitary of the catfish, *Clarias batrachus*: Molecular characterization and regulation by insulin. *The Journal of comparative neurology*, 530(11), 1743.

Zhang B, et al. (2021) Reconstruction of the Hypothalamo-Neurohypophysial System and Functional Dissection of Magnocellular Oxytocin Neurons in the Brain. *Neuron*, 109(2), 331.

Kawakami N, et al. (2021) Variation of pro-vasopressin processing in parvocellular and magnocellular neurons in the paraventricular nucleus of the hypothalamus: Evidence from the vasopressin-related glycopeptide copeptin. *The Journal of comparative neurology*, 529(7), 1372.

Otubo A, et al. (2020) Vasopressin gene products are colocalised with corticotrophin-releasing factor within neurosecretory vesicles in the external zone of the median eminence of the Japanese macaque monkey (*Macaca fuscata*). *Journal of neuroendocrinology*, 32(8), e12875.

DiBenedictis BT, et al. (2017) Quantitative mapping reveals age and sex differences in vasopressin, but not oxytocin, immunoreactivity in the rat social behavior neural network. *The Journal of comparative neurology*, 525(11), 2549.

de Kloet AD, et al. (2016) Angiotensin Type-2 Receptors Influence the Activity of Vasopressin Neurons in the Paraventricular Nucleus of the Hypothalamus in Male Mice. *Endocrinology*, 157(8), 3167.

Dimitrov E, et al. (2010) Tuberoinfundibular peptide of 39 residues modulates the mouse hypothalamic-pituitary-adrenal axis via paraventricular glutamatergic neurons. *The Journal of comparative neurology*, 518(21), 4375.

Kádár A, et al. (2010) Distribution of hypophysiotropic thyrotropin-releasing hormone (TRH)-synthesizing neurons in the hypothalamic paraventricular nucleus of the mouse. *The Journal of comparative neurology*, 518(19), 3948.

Belenky MA, et al. (2008) Heterogeneous expression of gamma-aminobutyric acid and gamma-aminobutyric acid-associated receptors and transporters in the rat suprachiasmatic nucleus. *The Journal of comparative neurology*, 506(4), 708.