

Resource Summary Report

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New York University Grossman School of Medicine Langone Health Cytometry and Cell Sorting Laboratory Core Facility

RRID:SCR_019179

Type: Tool

Proper Citation

New York University Grossman School of Medicine Langone Health Cytometry and Cell Sorting Laboratory Core Facility (RRID:SCR_019179)

Resource Information

URL: <https://med.nyu.edu/research/scientific-cores-shared-resources/cytometry-cell-sorting-laboratory>

Proper Citation: New York University Grossman School of Medicine Langone Health Cytometry and Cell Sorting Laboratory Core Facility (RRID:SCR_019179)

Description: Provides access to flow cytometry and cell sorting technologies and instruments. If your research requires cytometric analysis, instruments acquire optical measurements using different lasers to detect fluorophores with high level of precision.

Synonyms: Cytometry and Cell Sorting Laboratory

Resource Type: access service resource, core facility, service resource

Keywords: flow cytometry, cell sorting, cytometric analysis, ABRF

Funding: NCI P30CA016087

Resource Name: New York University Grossman School of Medicine Langone Health Cytometry and Cell Sorting Laboratory Core Facility

Resource ID: SCR_019179

Alternate IDs: ABRF_821

Alternate URLs: <https://coremarketplace.org/?FacilityID=821>

Record Creation Time: 20220129T080343+0000

Record Last Update: 20260912T200409+0000

Ratings and Alerts

No rating or validation information has been found for New York University Grossman School of Medicine Langone Health Cytometry and Cell Sorting Laboratory Core Facility.

No alerts have been found for New York University Grossman School of Medicine Langone Health Cytometry and Cell Sorting Laboratory Core Facility.

Data and Source Information

Source: [SciCrunch Registry](#)

Usage and Citation Metrics

We found 22 mentions in open access literature.

Listed below are recent publications. The full list is available at [SPARC SAWG](#) .

Damani-Yokota P, et al. (2026) Type 2 immune history imprints local training in nerve and airway associated interstitial macrophages (NAMS) for disease tolerance during lethal respiratory viral infection. Research square.

Drmi? D, et al. (2026) ZIP3 is essential for the encephalitogenic function of pTh17 cells by regulating intracellular zinc levels. Journal of trace elements in medicine and biology : organ of the Society for Minerals and Trace Elements (GMS), 93, 127819.

Noyer L, et al. (2025) ORAI1 mutation with mixed loss and gain of function properties causes immunodeficiency and HLH. Journal of human immunity, 1(4).

Nakamura R, et al. (2025) High dose methylprednisolone mediates YAP/TAZ-TEAD in vocal fold fibroblasts with macrophages. Scientific reports, 15(1), 11005.

Mudianto T, et al. (2025) Lymphatic constraint of germinal centers optimizes protective antibody responses. bioRxiv : the preprint server for biology.

Laskou M, et al. (2025) Platelets impair the resolution of inflammation in atherosclerotic plaques in insulin-resistant mice after lipid lowering. JCI insight, 10(21).

Pucella JN, et al. (2025) Tonic type I interferon signaling optimizes the antiviral function of plasmacytoid dendritic cells. Nature immunology, 26(11), 1946.

Coleman KE, et al. (2025) SLFN11 counteracts the RFW3-PRIMPOL DNA damage tolerance axis to restrain gapped DNA synthesis in response to replication stress. Nature communications, 16(1), 11029.

Newman AAC, et al. (2025) Ischemic Injury Drives Nascent Tumor Growth Via Accelerated Hematopoietic Aging. JACC. CardioOncology, 7(5), 559.

McDermott M, et al. (2025) Machine learning approach to single cell transcriptomic analysis of Sjogren's disease reveals altered activation states of B and T lymphocytes.

Journal of autoimmunity, 154, 103419.

Yeung ST, et al. (2025) Nerve- and airway-associated interstitial macrophages mitigate SARS-CoV-2 pathogenesis via type I interferon signaling. *Immunity*, 58(5), 1327.

Zhong L, et al. (2025) STIM1-mediated NFAT signaling synergizes with STAT1 to control T-bet expression and TH1 differentiation. *Nature immunology*, 26(3), 484.

Tao AY, et al. (2025) SLC7A5 regulates B cell metabolism and plasma cell differentiation independent of leucine transport. *Journal of immunology* (Baltimore, Md. : 1950).

Subhan BS, et al. (2025) Duo-nano exosome encapsulating hydrogel boosts wound healing across xenogenic and allogenic models. *Biomaterials*, 329, 123953.

Samper N, et al. (2024) Kir6.1, a component of an ATP-sensitive potassium channel, regulates natural killer cell development. *Frontiers in immunology*, 15, 1490250.

Nakamura R, et al. (2024) High-dose methylprednisolone mediates YAP/TAZ-TEAD in vocal fold fibroblasts with macrophages. *Research square*.

Barcia Durán JG, et al. (2024) Immune checkpoint landscape of human atherosclerosis and influence of cardiometabolic factors. *Nature cardiovascular research*, 3(12), 1482.

Cattle MA, et al. (2024) An enhanced Eco1 retron editor enables precision genome engineering in human cells from a single-copy integrated lentivirus. *bioRxiv : the preprint server for biology*.

Calderon A, et al. (2024) Chromatin accessibility and cell cycle progression are controlled by the HDAC-associated Sin3B protein in murine hematopoietic stem cells. *Epigenetics & chromatin*, 17(1), 2.

Samper N, et al. (2024) Kir6.1, a component of an ATP-sensitive potassium channel, regulates natural killer cell development. *bioRxiv : the preprint server for biology*.