Resource Summary Report

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Tetrahymena Stock Center

RRID:SCR_008362
Type: Tool

Proper Citation

Tetrahymena Stock Center (RRID:SCR_008362)

Resource Information

URL: https://tetrahymena.vet.cornell.edu/

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Description: Centralized repository and distribution site for variety of Tetrahymena strains and species. Maintains diverse array of wild type, mutant, and genetically engineered strains of T. thermophila, the most commonly used laboratory species, and variety of other species derived from both laboratory maintained stocks and wild isolates. All stocks are stored in liquid nitrogen to maintain genetic integrity and prevent senescence. In addition to providing worldwide access to strains currently in collection, TSC continually upgrades collection by accepting deposition of newly developed laboratory strains and well characterized wild isolates collected from clearly defined natural sites.

Abbreviations: TSC

Synonyms: Resource Center For Tetrahymena Thermophila

Resource Type: biomaterial supply resource, material resource

Keywords: RIN, Resource Information Network, eukaryota, genetics, biology, cell, cellular, culture, model, molecular, protozoan, repository, research, tetrahymena thermophila, RRID Community Authority

Funding: NIH Office of the Director P40 OD010964

Resource Name: Tetrahymena Stock Center

Resource ID: SCR_008362

Alternate IDs: nif-0000-25476

Old URLs: http://vivo.cornell.edu/individual/vivo/individual27605

Record Creation Time: 20220129T080247+0000

Record Last Update: 20260912T200237+0000

Ratings and Alerts

No rating or validation information has been found for Tetrahymena Stock Center.

No alerts have been found for Tetrahymena Stock Center.

Data and Source Information

Source: SciCrunch Registry

Usage and Citation Metrics

We found 89 mentions in open access literature.

Listed below are recent publications. The full list is available at SPARC SAWG .

Zhang Z, et al. (2026) Bromodomain protein IBD1 bridges histone acetylation and H2A.Z deposition to fine-tune transcription. Nucleic acids research, 54(4).

Cheng T, et al. (2025) Identification and characterization of the de novo methyltransferases for eukaryotic N6-methyladenine (6mA). Science advances, 11(20), eadq4623.

Ye F, et al. (2025) Comprehensive genome annotation of the model ciliate Tetrahymena thermophila by in-depth epigenetic and transcriptomic profiling. Nucleic acids research, 53(2).

Lei W, et al. (2025) Cystathionine ?-synthase TtCbs1 from Tetrahymena thermophila catalyzes the synthesis of CdS quantum dots for methyl orange decolorization. Applied and environmental microbiology, 91(10), e0125525.

Wang Y, et al. (2025) Dual modes of DNA N6-methyladenine maintenance by distinct methyltransferase complexes. Proceedings of the National Academy of Sciences of the United States of America, 122(3), e2413037121.

Duan L, et al. (2025) Methyl-dependent auto-regulation of the DNA N6-adenine methyltransferase AMT1 in the unicellular eukaryote Tetrahymena thermophila. Nucleic acids research, 53(3).

Zhang W, et al. (2025) Metabolomics' Change Under ?-Cypermethrin Stress and Detoxification Role of CYP5011A1 in Tetrahymena thermophila. Metabolites, 15(3).

Garg J, et al. (2025) Bromodomain proteins IBD1 and IBD2 link histone acetylation to SWR1- and INO80-mediated H2A.Z regulation in Tetrahymena. Epigenetics & chromatin, 18(1), 51.

Sheng Y, et al. (2024) Semiconservative transmission of DNA N 6-adenine methylation in a unicellular eukaryote. Genome research, 34(5), 740.

Cole ES, et al. (2024) The 'Janus A' gene encodes a polo-kinase whose loss creates a dorsal/ventral intracellular homeosis in the ciliate, Tetrahymena. bioRxiv : the preprint server for biology.

Ma Y, et al. (2024) Cip1, a CDK regulator, determines heterothallic mating or homothallic selfing in a protist. Proceedings of the National Academy of Sciences of the United States of America, 121(13), e2315531121.

Hao H, et al. (2023) Independent and Complementary Functions of Caf1b and Hir1 for Chromatin Assembly in Tetrahymena thermophila. Cells, 12(24).

Nabeel-Shah S, et al. (2023) Multilevel interrogation of H3.3 reveals a primordial role in transcription regulation. Epigenetics & chromatin, 16(1), 10.

Fukuda Y, et al. (2022) Snf2 Proteins Are Required to Generate Gamete Pronuclei in Tetrahymena thermophila. Microorganisms, 10(12).

Qiao Y, et al. (2022) Identification and utilization of a mutated 60S ribosomal subunit coding gene as an effective and cost-efficient selection marker for Tetrahymena genetic manipulation. International journal of biological macromolecules, 204, 1.

Lian Y, et al. (2022) Histone Chaperone Nrp1 Mutation Affects the Acetylation of H3K56 in Tetrahymena thermophila. Cells, 11(3).

Gotesman M, et al. (2022) Using a Hand-Held Gene Gun for Genetic Transformation of Tetrahymena thermophila. Methods in molecular biology (Clifton, N.J.), 2364, 349.

Lian Y, et al. (2021) The histone chaperone Nrp1 is required for chromatin stability and nuclear division in Tetrahymena thermophila. Epigenetics & chromatin, 14(1), 34.

Rao Q, et al. (2021) Structures of outer-arm dynein array on microtubule doublet reveal a motor coordination mechanism. Nature structural & molecular biology, 28(10), 799.

Liu Y, et al. (2021) An Optimized and Versatile Counter-Flow Centrifugal Elutriation Workflow to Obtain Synchronized Eukaryotic Cells. Frontiers in cell and developmental biology, 9, 664418.