

Resource Summary Report

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Louisiana State University in Shreveport Research Core Facility

RRID:SCR_024775

Type: Tool

Proper Citation

Louisiana State University in Shreveport Research Core Facility (RRID:SCR_024775)

Resource Information

URL: <https://research.lsuhs.edu/cores/research-core-facilities>

Proper Citation: Louisiana State University in Shreveport Research Core Facility (RRID:SCR_024775)

Description: Core provides access to research services, educational and training opportunities, and technologies including cellular metabolism, flow cytometry, fluorescence, confocal, super resolution confocal microscopy, next generation sequencing, and real-time PCR. Provides assistance with all aspects of these technologies, including experimental design, protocol optimization, instrument operation, and data analysis.

Abbreviations: RCF

Synonyms: Research Core Facility

Resource Type: core facility, access service resource, service resource

Keywords: ABRF, cellular metabolism, flow cytometry, fluorescence, confocal microscopy, super resolution confocal microscopy, next generation sequencing, RT PCR,

Availability: Open

Resource Name: Louisiana State University in Shreveport Research Core Facility

Resource ID: SCR_024775

Alternate IDs: ABRF_2566

Alternate URLs: <https://coremarketplace.org/?FacilityID=2566&citation=1>

Record Creation Time: 20231212T050231+0000

Ratings and Alerts

No rating or validation information has been found for Louisiana State University in Shreveport Research Core Facility.

No alerts have been found for Louisiana State University in Shreveport Research Core Facility.

Data and Source Information

Source: [SciCrunch Registry](#)

Usage and Citation Metrics

We found 23 mentions in open access literature.

Listed below are recent publications. The full list is available at [SPARC SAWG](#) .

Arias F, et al. (2026) Targeting Oxalate Production by Combining Enzyme Inhibition and Proteolysis Activation: A Novel Therapeutic Approach for Primary Hyperoxaluria Type 1. Journal of medicinal chemistry.

First NJ, et al. (2026) Protocol for visualization and quantitative analysis of murine lung immunological architecture using iBALT-UNMAP following bacterial challenge. STAR protocols, 7(2), 104472.

Word C, et al. (2026) Expanded protocadherin-1 usage reveals a broader hantavirus entry landscape. bioRxiv : the preprint server for biology.

Dhungel N, et al. (2026) Direct interaction between cancer cells and fibroblasts promotes early chemoresistance to standard-of-care drug therapy in small cell lung cancer (SCLC). bioRxiv : the preprint server for biology.

Parrish KM, et al. (2025) The Bordetella type III secretion system effector BteA targets host eosinophil-epithelial signaling to promote IL-1Ra expression and persistence. Communications biology, 8(1), 1484.

Mohajan S, et al. (2025) Nuclear basket proteins Nup2 and Mlp1 drive heat shock-induced 3D genome restructuring downstream of transcriptional activation. The Journal of biological chemistry, 301(9), 110568.

Mohajan S, et al. (2025) Nuclear basket proteins Mlp1 and Nup2 drive heat shock-induced 3D genome restructuring. bioRxiv : the preprint server for biology.

Lee M, et al. (2025) Methamphetamine-Induced Loss of Syndecan-1 and Retinal Endothelial Integrity via the TAAR-1/MMP-9 Pathway. Pathophysiology : the official journal of the International Society for Pathophysiology, 32(3).

Ben Dhaou C, et al. (2025) Integrin-specific signaling drives ER stress-dependent atherogenic endothelial activation. *Redox biology*, 88, 103911.

Zong X, et al. (2025) Clemastine ameliorates cognitive deficits in an experimental rat model of chronic cerebral hypoperfusion. *Journal of cerebral blood flow and metabolism : official journal of the International Society of Cerebral Blood Flow and Metabolism*, 271678X251382874.

Ogundepo O, et al. (2025) Studies on the functionality of the TC-NER ERCC6-M1097V protein variant frequently found in Louisiana patients with PCa upon UV damage. *Frontiers in oncology*, 15, 1679379.

Keiffer TR, et al. (2025) HPV16 entry requires dynein for minus-end transport and utilizes kinesin Kif11 for plus-end transport along microtubules during mitosis. *Journal of virology*, 99(1), e0093724.

Lee M, et al. (2025) Retinal Angiogenesis in Methamphetamine Self-Administration Rats. *Investigative ophthalmology & visual science*, 66(9), 8.

Norris MJ, et al. (2025) The GATE glycoprotein complex enhances human cytomegalovirus entry in endothelial cells. *Nature microbiology*, 10(7), 1605.

Kaur H, et al. (2025) Neutrophil Integrin $\alpha 9$ Impairs Efferocytosis and Worsens Long-Term Recovery After Subarachnoid Hemorrhage. *Arteriosclerosis, thrombosis, and vascular biology*.

Pearson-Gallion B, et al. (2025) Fibronectin-dependent integrin signaling drives EphA2 expression in vascular smooth muscle cells. *American journal of physiology. Cell physiology*, 328(5), C1623.

Dhaou CB, et al. (2025) Integrin-Specific Signaling Drives ER Stress-Dependent Atherogenic Endothelial Activation. *bioRxiv : the preprint server for biology*.

Aishwarya R, et al. (2024) Diastolic dysfunction in Alzheimer's disease model mice is associated with A β -amyloid aggregate formation and mitochondrial dysfunction. *Scientific reports*, 14(1), 16715.

Norris MJ, et al. (2024) A noncanonical glycoprotein H complex enhances cytomegalovirus entry. *bioRxiv : the preprint server for biology*.

Pandey N, et al. (2024) Interactions between integrin $\alpha 9 \beta 1$ and VCAM-1 promote neutrophil hyperactivation and mediate poststroke DVT. *Blood advances*, 8(9), 2104.