

Resource Summary Report

Generated by [SPARC SAWG](#) on Aug 9, 2026

Artemis: Genome Browser and Annotation Tool

RRID:SCR_004267

Type: Tool

Proper Citation

Artemis: Genome Browser and Annotation Tool (RRID:SCR_004267)

Resource Information

URL: <http://www.sanger.ac.uk/resources/software/artemis/>

Proper Citation: Artemis: Genome Browser and Annotation Tool (RRID:SCR_004267)

Description: THIS RESOURCE IS NO LONGER IN SERVICE. Documented on February 28, 2023. Free genome browser and annotation tool that allows visualization of sequence features, next generation data and the results of analyses within the context of the sequence, and also its six-frame translation. Artemis is free software and is distributed under the terms of the GNU General Public License. Artemis is written in Java, and is available for UNIX, Macintosh and Windows systems. It can read EMBL and GENBANK database entries or sequence in FASTA, indexed FASTA or raw format. Other sequence features can be in EMBL, GENBANK or GFF format.

Abbreviations: Artemis

Resource Type: software resource

Defining Citation: [PMID:11120685](#), [DOI:10.1093/bioinformatics/btr703](#)

Keywords: training tool, genome browser, gene annotation, java, bio.tools

Funding: Wellcome Trust

Availability: THIS RESOURCE IS NO LONGER IN SERVICE

Resource Name: Artemis: Genome Browser and Annotation Tool

Resource ID: SCR_004267

Alternate IDs: nlx_28554, OMICS_00903, biotools:artemis

Alternate URLs: <https://bio.tools/artemis>, <https://sources.debian.org/src/art-nextgen-simulation-tools/>

Record Creation Time: 20220129T080223+0000

Record Last Update: 20260808T185814+0000

Ratings and Alerts

No rating or validation information has been found for Artemis: Genome Browser and Annotation Tool.

No alerts have been found for Artemis: Genome Browser and Annotation Tool.

Data and Source Information

Source: [SciCrunch Registry](#)

Usage and Citation Metrics

We found 422 mentions in open access literature.

Listed below are recent publications. The full list is available at [SPARC SAWG](#) .

Withatanung P, et al. (2026) Genomic diversity and clade clustering of Burkholderia pseudomallei and B. thailandensis prophages with soil-derived phages. iScience, 29(2), 114658.

Yu L, et al. (2025) Parasitic structure defect blights sustainability of cobalt-free single crystalline cathodes. Nature communications, 16(1), 434.

Zhang Z, et al. (2025) Breaking linear scaling relationships in oxygen evolution via dynamic structural regulation of active sites. Nature communications, 16(1), 1301.

Bienenmann RLM, et al. (2025) Chloride, Alkoxide, or Silicon: The Bridging Ligand Dictates the Spin State in Dicobalt Expanded Pincer Complexes. Organometallics, 44(1), 94.

Yin J, et al. (2025) Unconventional hexagonal open Prussian blue analog structures. Nature communications, 16(1), 370.

Li H, et al. (2025) Chemical looping synthesis of amines from N₂ via iron nitride as a mediator. Nature communications, 16(1), 257.

Yacoub E, et al. (2025) A sweeping view of avian mycoplasmas biology drawn from comparative genomic analyses. BMC genomics, 26(1), 24.

Haenni M, et al. (2025) Distinct molecular epidemiology of resistances to extended-spectrum cephalosporins and carbapenems in Enterobacter hormaechei in cats and dogs versus horses in France. The Journal of antimicrobial chemotherapy, 80(2), 567.

Wang K, et al. (2025) Visible-light-driven CO₂ photoreduction over atomically strained indium sites in ambient air. Nature communications, 16(1), 2094.

Zakharzhevskaya NB, et al. (2025) Proteogenomic annotation of T6SS components identified in *Bacteroides fragilis* secretome. *Frontiers in microbiology*, 16, 1495971.

Santini S, et al. (2025) Pacmanvirus isolated from the Lost City hydrothermal field extends the concept of transpoviron beyond the family Mimiviridae. *The ISME journal*, 19(1).

Zhang K, et al. (2025) Thickness-dependent polaron crossover in tellurene. *Science advances*, 11(2), eads4763.

Yan Y, et al. (2025) General synthesis of neighboring dual-atomic sites with a specific pre-designed distance via an interfacial-fixing strategy. *Nature communications*, 16(1), 334.

Wang T, et al. (2025) Bioinspired artificial antioxidantases for efficient redox homeostasis and maxillofacial bone regeneration. *Nature communications*, 16(1), 856.

Zhang T, et al. (2025) Atomically thin high-entropy oxides via naked metal ion self-assembly for proton exchange membrane electrolysis. *Nature communications*, 16(1), 1037.

White RT, et al. (2025) The rapid detection of a neonatal unit outbreak of a wild-type *Klebsiella variicola* using decentralized Oxford Nanopore sequencing. *Antimicrobial resistance and infection control*, 14(1), 6.

Maul JE, et al. (2025) Genomic and mutational analysis of *Pseudomonas syringae* pv. *tagetis* EB037 pathogenicity on sunflower. *BMC microbiology*, 25(1), 43.

Xie Z, et al. (2024) Well-defined diatomic catalysis for photosynthesis of C₂H₄ from CO₂. *Nature communications*, 15(1), 2422.

Butburee T, et al. (2024) General Pyrolysis for High-Loading Transition Metal Single Atoms on 2D-Nitro-Oxygeneous Carbon as Efficient ORR Electrocatalysts. *ACS applied materials & interfaces*, 16(8), 10227.

Niu W, et al. (2024) High-efficiency C₃ electrosynthesis on a lattice-strain-stabilized nitrogen-doped Cu surface. *Nature communications*, 15(1), 7070.