Resource Summary Report

Generated by PRECISE-TBI on Sep 8, 2026

Emory University Emory Glycomics and Molecular Interactions Core Facility

RRID:SCR_023524
Type: Tool

Proper Citation

Emory University Emory Glycomics and Molecular Interactions Core Facility (RRID:SCR_023524)

Resource Information

URL: https://www.cores.emory.edu/egmic/

Proper Citation: Emory University Emory Glycomics and Molecular Interactions Core Facility (RRID:SCR_023524)

Description: Provides access to methods and technology in area of glycomics and molecular interactions.Provides individual consultation with investigators to address questions regarding glycosylation, the most common posttranslational modification, and protein glycan interactions.Provides access to mass spectrometry and glycan microarray technologies to help understand functions of glycans and glycan binding proteins.

Abbreviations: EGMIC

Synonyms: Emory University Emory Glycomics and Molecular Interactions Core (EGMIC), Emory Glycomics and Molecular Interactions Core (EGMIC)

Resource Type: access service resource, core facility, service resource

Keywords: USEDit, ABRF, glycomics and molecular interactions services,

Availability: Open

Resource Name: Emory University Emory Glycomics and Molecular Interactions Core Facility

Resource ID: SCR_023524

Alternate IDs: ABRF_1740

Alternate URLs: https://coremarketplace.org/?FacilityID=1740&citation=1

Record Creation Time: 20230503T050210+0000

Record Last Update: 20260905T133435+0000

Ratings and Alerts

No rating or validation information has been found for Emory University Emory Glycomics and Molecular Interactions Core Facility.

No alerts have been found for Emory University Emory Glycomics and Molecular Interactions Core Facility.

Data and Source Information

Source: SciCrunch Registry

Usage and Citation Metrics

We found 10 mentions in open access literature.

Listed below are recent publications. The full list is available at PRECISE-TBI .

Galindo AV, et al. (2026) Cooperative Aldehyde Chemistry Maps an Orthogonal Lysine Reactivity Landscape. Journal of the American Chemical Society, 148(14), 15125.

Emenike B, et al. (2026) Unlocking the Silent Proteome: Chemoselective Asn/Gln Activation for Multidimensional Protein Diversification. Journal of the American Chemical Society, 148(11), 11974.

Kumari P, et al. (2026) Neo-Cysteine Molecular Glues for Targeting Mutated SMAD4 Protein. Angewandte Chemie (International ed. in English), 65(14), e12791.

Zhang Q, et al. (2026) Preparative separation of ?2,3- and ?2,6-sialylated N-glycans via linkage-specific charge modulation. Carbohydrate research, 563, 109867.

Paikin ZE, et al. (2026) Reprogramming the lipid peroxidation product 4-ONE as a chemoselective cleavable crosslinker. Nature communications.

Zhang Q, et al. (2025) Streamlined gram-scale natural N-glycan production using reversible tagging after oxidative release of natural glycans. Communications chemistry, 8(1), 103.

Lin B, et al. (2025) Testing the Luedemann hypothesis: the discovery of novel antimicrobials from slow-growing microbes from nutrient-limited environments. mSphere, e0036725.

Zhang H, et al. (2025) Sample-sparing multiplexed antibody Fc biomarker discovery using a reconfigurable integrated microfluidic platform. Lab on a chip, 25(12), 2828.

Zhang Q, et al. (2025) Oxidative Release of Natural Glycans (ORNG): Preparation of Truncated N-glycans for Engineering Glycoproteins. Chemistry (Weinheim an der Bergstrasse, Germany), 31(32), e202501020.

Zhang Q, et al. (2024) Oxidative Release of Natural Glycans: Unraveling the Mechanism for Rapid N-Glycan Glycomics Analysis. Analytical chemistry, 96(42), 16750.