

# Resource Summary Report

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## A plasmid Editor

RRID:SCR\_014266

Type: Tool

### Proper Citation

A plasmid Editor (RRID:SCR\_014266)

### Resource Information

**URL:** <http://biologylabs.utah.edu/jorgensen/wayned/ape/>

**Proper Citation:** A plasmid Editor (RRID:SCR\_014266)

**Description:** Software tool for plasmid and sequence editing, annotating and drawing plasmid sequences. Used to view circular or linear maps of DNA sequences. Users can perform virtual digests whereby they select predefined DNA ladder, or specify their own, and visualize theoretical DNA fragments. Used to highlight restriction sites in editing window, accurately reflect Dam/Dcm blocking of enzyme sites, highlighting and drawing graphic maps using feature annotations from genbank and embl files, highlighting text using pre-defined and custom feature libraries, and directly BLASTing selected sequence at NCBI or Wormbase. Runs across Windows, OS X, and Linux/Unix.

**Abbreviations:** ApE

**Synonyms:** A plasmid Editor

**Resource Type:** data analysis software, software application, software resource, sequence analysis software, data processing software, standalone software

**Keywords:** Plasmid, editing, sequence, annotating, drawing, restriction, site, enzyme, map, DNA, fragment

**Availability:** Free, Available for download, Freely available

**Resource Name:** A plasmid Editor

**Resource ID:** SCR\_014266

**Alternate URLs:** <https://jorgensen.biology.utah.edu/wayned/ape/>

**Old URLs:** <http://ape-a-plasmid-editor.wikispaces.com>

**Record Creation Time:** 20220129T080319+0000

## Ratings and Alerts

No rating or validation information has been found for A plasmid Editor.

No alerts have been found for A plasmid Editor.

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## Data and Source Information

**Source:** [SciCrunch Registry](#)

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## Usage and Citation Metrics

We found 93 mentions in open access literature.

**Listed below are recent publications.** The full list is available at [PRECISE-TBI](#) .

Horsley N, et al. (2026) CrisprBuildr: an open-source application for CRISPR-mediated genome engineering in *Drosophila melanogaster*. *G3* (Bethesda, Md.), 16(1).

Jin EJ, et al. (2025) The BEN domain protein LIN-14 coordinates neuromuscular positioning during epidermal maturation. *iScience*, 28(1), 111577.

Schiffmaier J, et al. (2025) Establishment of human periodontal ligament cell lines with ALPL mutations to mimic dental aspects of hypophosphatasia. *Frontiers in cell and developmental biology*, 13, 1572571.

Kengne-Ouafo JA, et al. (2025) Plasmodium falciparum Subtilisin-like Domain-Containing Protein (PfSDP), a Cross-Stage Antigen, Elicits Short-Lived Antibody Response Following Natural Infection with Plasmodium falciparum. *Cells*, 14(15).

Nie J, et al. (2025) T2T-CHM13 versus hg38: accurate identification of immunoglobulin isotypes from scRNA-seq requires a genome reference matched for ancestry. *NAR genomics and bioinformatics*, 7(2), lqaf074.

Imai Y, et al. (2025) Real-time imaging of blood coagulation and angiogenesis during development in a zebrafish model of type I antithrombin deficiency. *Scientific reports*, 15(1), 18538.

Horsley N, et al. (2025) CrisprBuildr: an open-source application for CRISPR-mediated genome engineering in *Drosophila melanogaster*. *bioRxiv : the preprint server for biology*.

Brinkert P, et al. (2025) The actin nucleation promoting factor WASH facilitates clathrin-independent endocytosis of human papillomaviruses. *EMBO reports*, 26(22), 5533.

Parker DM, et al. (2025) Protocol for determining the contribution of protein and RNA to condensate organization by permeabilizing and enzyme-treating live U-2 OS cells. *STAR protocols*, 6(2), 103744.

Banerjee N, et al. (2025) Protocol to generate stable knockout lines in the human-parasitic nematode *Strongyloides stercoralis*. STAR protocols, 6(4), 104201.

Thakur RS, et al. (2025) PDZD8 promotes autophagy at ER-lysosome membrane contact sites to regulate activity-dependent synaptic growth. Cell reports, 44(4), 115483.

Regan SB, et al. (2025) Megabase-scale loss of heterozygosity provoked by CRISPR-Cas9 DNA double-strand breaks. Molecular cell, 85(22), 4119.

Purice MD, et al. (2025) Molecular profiling of adult *C. elegans* glia across sexes by single-nuclear RNA-seq. Developmental cell.

Modaffari D, et al. (2024) Improved gene editing and fluorescent-protein tagging in *Aspergillus nidulans* using a Golden Gate-based CRISPR-Cas9 plasmid system. Wellcome open research, 9, 602.

Noguchi Y, et al. (2024) In vivo CRISPR screening directly targeting testicular cells. Cell genomics, 4(3), 100510.

Durante-Rodríguez G, et al. (2024) Characterization of a MHYT domain-coupled transcriptional regulator that responds to carbon monoxide. Nucleic acids research, 52(15), 8849.

Chen TQ, et al. (2024) Comparative Mitogenomics Provides Valuable Insights for the Phylogeny and New DNA Barcodes of *Ganoderma*. Journal of fungi (Basel, Switzerland), 10(11).

Kitashin I, et al. (2024) Complete genome sequences of feline astroviruses from asymptomatic stray cats in Japan. Microbiology resource announcements, 13(11), e0071224.

Kojima L, et al. (2024) Optimization of AAV vectors for transactivator-regulated enhanced gene expression within targeted neuronal populations. iScience, 27(6), 109878.

Geraud M, et al. (2024) TDP1 mutation causing SCAN1 neurodegenerative syndrome hampers the repair of transcriptional DNA double-strand breaks. Cell reports, 43(5), 114214.