Resource Summary Report

Generated by nidm-terms on Sep 21, 2026

University of California San Francisco Parnassus Flow Cytometry Core Facility

RRID:SCR_018206
Type: Tool

Proper Citation

University of California San Francisco Parnassus Flow Cytometry Core Facility (RRID:SCR_018206)

Resource Information

URL: https://flow.ucsf.edu/

Proper Citation: University of California San Francisco Parnassus Flow Cytometry Core Facility (RRID:SCR_018206)

Description: Core assists investigators whose research requires molecular marker characterization of cells in suspension as well as isolation of cells based on those markers. Advanced cell sorting and cytometric analyses by Flow or Mass Cytometry are provided.

Abbreviations: PFCC

Synonyms: University of California San Francisco Diabetes Research Center Cytometry and Cell Sorting Core, , University of California San Francisco Diabetes Research Center Parnassus Flow Cytometry CoLab, UCSF Parnassus Flow CoLab

Resource Type: access service resource, core facility, service resource

Keywords: Molecular marker, cell characterization, cell in suspension, cell isolation, cell sorting, flow cytometry, mass cytometry, cytometric analysis, core facility, ABRF

Funding: NIDDK P30DK063720

Availability: Restricted

Resource Name: University of California San Francisco Parnassus Flow Cytometry Core Facility

Resource ID: SCR_018206

Alternate IDs: ABRF_1007, SCR_015105

Alternate URLs: https://coremarketplace.org?FacilityID=1007

Record Creation Time: 20220129T080339+0000

Record Last Update: 20260919T195935+0000

Ratings and Alerts

No rating or validation information has been found for University of California San Francisco Parnassus Flow Cytometry Core Facility.

No alerts have been found for University of California San Francisco Parnassus Flow Cytometry Core Facility.

Data and Source Information

Source: SciCrunch Registry

Usage and Citation Metrics

We found 174 mentions in open access literature.

Listed below are recent publications. The full list is available at nidm-terms .

Nyberg WA, et al. (2026) In vivo site-specific engineering to reprogram T cells. Nature, 652(8110), 712.

Superville DA, et al. (2026) Tumor Heterogeneity Induces Pro- and Anti-metastatic Myeloid Cell Phenotypes in Breast Cancer Lung Metastasis. bioRxiv : the preprint server for biology.

Lee EY, et al. (2026) Computational discovery of precision therapeutics for hidradenitis suppurativa. bioRxiv : the preprint server for biology.

Levan J, et al. (2026) Congenital cytomegalovirus infection drives oligoclonal expansion of cytotoxic ?? T cells from early fetal progenitors. Journal of immunology (Baltimore, Md. : 1950), 215(6).

Berleant JD, et al. (2026) Enabling global-scale nucleic acid repositories through versatile, scalable biochemical selection from room-temperature archives. Nature communications, 17(1).

Fan AC, et al. (2026) Submicrometre sampling of living cells by macrophages. Nature, 654(8118), 495.

Moss T, et al. (2026) Hsp70 is phosphorylated in a conserved response to DNA damage and contributes to cell cycle control. eLife, 15.

Tamaki CM, et al. (2026) Gain-of-function mutation in SKAP2 leads to type 1 diabetes and broader autoimmunity through hyperactive integrin signaling in myeloid cells. bioRxiv : the

preprint server for biology.

Tharp CR, et al. (2026) Biophysical trade-offs in antibody evolution are resolved by conformation-mediated epistasis. bioRxiv : the preprint server for biology.

Sbierski-Kind J, et al. (2026) Type 2 lymphocytes restrict type 3 lymphocytes during liver fibrosis and colocalize in fibroblast niches. Science advances, 12(11), eaea6805.

Dwyer LR, et al. (2026) CD38 expression by neonatal human naive CD4+ T cells shapes their distinct metabolic and tolerogenic properties. The Journal of clinical investigation, 136(8).

Trivedi A, et al. (2026) A dried platelet-derived biologic for blood-brain barrier repair and hemorrhage control after TBI in mice. Blood, 147(26), 3231.

Berquez M, et al. (2026) A multi-subunit autophagic capture complex facilitates degradation of ER-stalled MHC class I in pancreatic cancer. Molecular cell, 86(11), 2141.

Sin JH, et al. (2026) Foxp1 controls Fezf2-dependent gene expression, mimetic mTEC diversity, and thymic central tolerance. Journal of immunology (Baltimore, Md. : 1950), 215(6).

Sin JH, et al. (2026) Runx3 instructs Aire+ mTEC development, TSA gene expression, and central tolerance. Journal of autoimmunity, 161, 103573.

Shen F, et al. (2026) Capsule modulation enhances immunity and delays resistance in MDR Acinetobacter baumannii. iScience, 29(6), 115761.

Sin JH, et al. (2026) Thymic alveolar type 2 epithelial mimetic cells revealed by RUNX1 deficiency. Nature immunology, 27(7), 1462.

Babdor J, et al. (2026) Immune-microbiome coordination defines interferon setpoints in healthy humans. Cell, 189(10), 3144.

Dhariwala MO, et al. (2026) Commensal-myeloid crosstalk in neonatal skin regulates interleukin-1 signaling and cutaneous type 17 inflammation. Immunity, 59(2), 403.

Umhoefer JM, et al. (2026) FOXP3 expression depends on cell-type-specific cis-regulatory elements and transcription factor circuitry. Immunity, 59(1), 129.