Resource Summary Report

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Virginia University School of Medicine Advanced Microscopy Core Facility

RRID:SCR_018736
Type: Tool

Proper Citation

Virginia University School of Medicine Advanced Microscopy Core Facility (RRID:SCR_018736)

Resource Information

URL: https://med.virginia.edu/advanced-microscopy-facility/

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Description: Advanced Microscopy Facility is microscopy service and shared instrumentation facility in University of Virginia biomedical research community. Pprovides expertise and instrument access for electron and light microscopy imaging technologies as well as image analysis tools and services for basic scientists and clinical researchers within University of Virginia and across Commonwealth.

Abbreviations: AMF

Synonyms: University of Virginia UVA Advanced Microscopy Facility, UVA Advanced Microscopy Facility

Resource Type: access service resource, core facility, service resource

Keywords: USEDit, microscopy, electron microscopy, light microscopy, imaging technology, image analysis, training, sample preparation, consultation, ABRF, ABRF

Availability: Open

Resource Name: Virginia University School of Medicine Advanced Microscopy Core Facility

Resource ID: SCR_018736

Alternate IDs: ABRF_1018, SCR_018744

Alternate URLs: https://coremarketplace.org/?FacilityID=1018, https://med.virginia.edu/advanced-microscopy-facility/services/

Record Creation Time: 20220129T080341+0000

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Ratings and Alerts

No rating or validation information has been found for Virginia University School of Medicine Advanced Microscopy Core Facility.

No alerts have been found for Virginia University School of Medicine Advanced Microscopy Core Facility.

Data and Source Information

Source: SciCrunch Registry

Usage and Citation Metrics

We found 19 mentions in open access literature.

Listed below are recent publications. The full list is available at nidm-terms .

Aragon JW, et al. (2025) Emergence of endothelial subtypes and role of cell cycle control in arterial-venous specification during embryonic vascular development. Cell reports, 44(10), 116368.

Leonard-Duke J, et al. (2025) Mechanically Tunable Poly(Ethylene Glycol) Diacrylate Hydrogels Reveal Stiffness-Related Impairments in Capillary Sprouting in Experimental Lung Fibrosis. Microcirculation (New York, N.Y. : 1994), 32(5), e70018.

Yadav N, et al. (2025) Synergistic activity of simvastatin and irinotecan chemotherapy against glioblastoma converges on TGF-?? signaling. Journal of neuro-oncology, 174(3), 621.

Daboin C, et al. (2025) Mechanical maturation of human dermal fibroblast-laden microporous annealed particle scaffolds during long-term in vitro culture. Acta biomaterialia, 207, 353.

Rao VG, et al. (2025) Mechanisms of cilia regeneration in Xenopus multiciliated epithelium in vivo. EMBO reports, 26(8), 2192.

Barbeau MC, et al. (2025) The kinase ERK plays a conserved dominant role in the heterogeneity of epithelial-mesenchymal transition in pancreatic cancer cells. Science signaling, 18(899), eads7002.

Zatorski JM, et al. (2024) Measurement of Covalent Bond Formation in Light-Curing Hydrogels Predicts Physical Stability under Flow. Analytical chemistry, 96(50), 19880.

Yamaguchi M, et al. (2024) Transformation of the Kidney into a Pathological Neuro-Immune-Endocrine Organ. Circulation research, 135(10), 1025.

Zeng J, et al. (2024) Protocol for integrating tissue clearing and light-sheet imaging to analyze cancer initiation in mosaic analysis with double markers mouse models. STAR protocols, 5(2), 103092.

Cox BP, et al. (2024) Local, Quantitative Morphometry of Fibroproliferative Lung Injury Using Laminin. American journal of respiratory cell and molecular biology, 71(1), 23.

Zhao XY, et al. (2024) iNOS is necessary for GBP-mediated T. gondii clearance in murine macrophages via vacuole nitration and intravacuolar network collapse. Nature communications, 15(1), 2698.

Zatorski JM, et al. (2024) Measurement of covalent bond formation in light-curing hydrogels predicts physical stability under flow. bioRxiv : the preprint server for biology.

Medrano S, et al. (2024) An efficient inducible model for the control of gene expression in renin cells. American journal of physiology. Renal physiology, 327(3), F489.

Przanowski P, et al. (2023) ANKLE1 cleaves mitochondrial DNA and contributes to cancer risk by promoting apoptosis resistance and metabolic dysregulation. Communications biology, 6(1), 231.

Zhao XY, et al. (2023) Inducible nitric oxide synthase (iNOS) is necessary for GBP-mediated T. gondii restriction in murine macrophages via vacuole nitration and intravacuolar network collapse. bioRxiv : the preprint server for biology.

Cox BP, et al. (2023) Local, Quantitative Morphometry of Fibroproliferative Lung Injury using Laminin. bioRxiv : the preprint server for biology.

Karanja F, et al. (2022) Ecdysone exerts biphasic control of regenerative signaling, coordinating the completion of regeneration with developmental progression. Proceedings of the National Academy of Sciences of the United States of America, 119(5).

DaCrema D, et al. (2021) Ecdysone regulates the Drosophila imaginal disc epithelial barrier, determining the length of regeneration checkpoint delay. Development (Cambridge, England), 148(6).

Patterson L, et al. (2020) Glucosylceramide production maintains colon integrity in response to Bacteroides fragilis toxin-induced colon epithelial cell signaling. FASEB journal : official publication of the Federation of American Societies for Experimental Biology, 34(12), 15922.