Resource Summary Report

Generated by nidm-terms on Aug 9, 2026

pcDNA-dCas9-VP64

RRID:Addgene_47107
Type: Plasmid

Proper Citation

RRID:Addgene_47107

Plasmid Information

URL: http://www.addgene.org/47107

Proper Citation: RRID:Addgene_47107

Insert Name: dCas9-VP64

Organism: S. Pyogenes

Bacterial Resistance: Ampicillin

Defining Citation: PMID:23892895

Vector Backbone Description: Backbone Marker:Invitrogen; Vector Backbone:pcDNA3.1; Vector Types:Mammalian Expression, CRISPR; Bacterial Resistance:Ampicillin

Plasmid Name: pcDNA-dCas9-VP64

Relevant Mutation: D10A, H840A (catalytically inactive)

Record Creation Time: 20220422T222242+0000

Record Last Update: 20260808T081728+0000

Ratings and Alerts

No rating or validation information has been found for pcDNA-dCas9-VP64.

No alerts have been found for pcDNA-dCas9-VP64.

Data and Source Information

Source: Addgene

Usage and Citation Metrics

We found 23 mentions in open access literature.

Listed below are recent publications. The full list is available at nidm-terms .

Pelea O, et al. (2025) Specific modulation of CRISPR transcriptional activators through RNA-sensing guide RNAs in mammalian cells and zebrafish embryos. eLife, 12.

Bamunuarachchi G, et al. (2025) Tankyrases positively regulate influenza A virus replication via type I interferon response. Journal of virology, 99(10), e0129825.

Ohtani H, et al. (2024) Efficient activation of hundreds of LTR12C elements reveals cis-regulatory function determined by distinct epigenetic mechanisms. Nucleic acids research, 52(14), 8205.

Huang H, et al. (2023) Epigenome editing based on CRISPR/dCas9p300 facilitates transdifferentiation of human fibroblasts into Leydig-like cells. Experimental cell research, 425(2), 113551.

Beyersdorf JP, et al. (2022) Robust, Durable Gene Activation In Vivo via mRNA-Encoded Activators. ACS nano, 16(4), 5660.

Rappaport JA, et al. (2022) A ?-Catenin-TCF-Sensitive Locus Control Region Mediates GUCY2C Ligand Loss in Colorectal Cancer. Cellular and molecular gastroenterology and hepatology, 13(4), 1276.

Ren C, et al. (2022) Highly efficient activation of endogenous gene in grape using CRISPR/dCas9-based transcriptional activators. Horticulture research, 9.

Weuring WJ, et al. (2021) CRISPRa-Mediated Upregulation of scn1laa During Early Development Causes Epileptiform Activity and dCas9-Associated Toxicity. The CRISPR journal, 4(4), 575.

Mózsik L, et al. (2021) Modular Synthetic Biology Toolkit for Filamentous Fungi. ACS synthetic biology, 10(11), 2850.

Jain S, et al. (2021) TALEN outperforms Cas9 in editing heterochromatin target sites. Nature communications, 12(1), 606.

Gamboa L, et al. (2020) Heat-Triggered Remote Control of CRISPR-dCas9 for Tunable Transcriptional Modulation. ACS chemical biology, 15(2), 533.

Polkoff KM, et al. (2020) In Vitro Validation of Transgene Expression in Gene-Edited Pigs Using CRISPR Transcriptional Activators. The CRISPR journal, 3(5), 409.

Matharu N, et al. (2019) CRISPR-mediated activation of a promoter or enhancer rescues obesity caused by haploinsufficiency. Science (New York, N.Y.), 363(6424).

Haenfler JM, et al. (2018) Targeted Reactivation of FMR1 Transcription in Fragile X Syndrome Embryonic Stem Cells. Frontiers in molecular neuroscience, 11, 282.

Pathak GP, et al. (2017) Bidirectional approaches for optogenetic regulation of gene expression in mammalian cells using Arabidopsis cryptochrome 2. Nucleic acids research, 45(20), e167.

Brown A, et al. (2017) Targeted Gene Activation Using RNA-Guided Nucleases. Methods in molecular biology (Clifton, N.J.), 1468, 235.

Bao Z, et al. (2017) Orthogonal Genetic Regulation in Human Cells Using Chemically Induced CRISPR/Cas9 Activators. ACS synthetic biology, 6(4), 686.

Jusiak B, et al. (2016) Engineering Synthetic Gene Circuits in Living Cells with CRISPR Technology. Trends in biotechnology, 34(7), 535.

Ji H, et al. (2016) Specific Reactivation of Latent HIV-1 by dCas9-SunTag-VP64-mediated Guide RNA Targeting the HIV-1 Promoter. Molecular therapy : the journal of the American Society of Gene Therapy, 24(3), 508.

Garcia-Bloj B, et al. (2016) Waking up dormant tumor suppressor genes with zinc fingers, TALEs and the CRISPR/dCas9 system. Oncotarget, 7(37), 60535.