

Resource Summary Report

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Tulane University TNBRC Confocal Microscopy and Molecular Pathology Core Facility

RRID:SCR_024613

Type: Tool

Proper Citation

Tulane University TNBRC Confocal Microscopy and Molecular Pathology Core Facility
(RRID:SCR_024613)

Resource Information

URL: <https://tnprc.tulane.edu/confocal-microscopy-and-molecular-pathology-core>

Proper Citation: Tulane University TNBRC Confocal Microscopy and Molecular Pathology Core Facility (RRID:SCR_024613)

Description: Core provides research support and training with molecular pathology skills, confocal microscopy, image analysis, and multicolor immunohistochemistry and in-situ hybridization techniques for internal and affiliate scientists. Services include assistance with experimental design, antibody selection, molecular probe selection, fixation and staining protocols, operating fluorescent and confocal microscopes, acquiring and storing imaging data, interpretation of results, and generation of publication-quality images. Image analysis services include assistance with capturing images for evaluation and with use of various image analysis software.

Abbreviations: CMMPC

Synonyms: , TNBRC Confocal Microscopy and Molecular Pathology Core, Tulane University TNBRC Confocal Microscopy and Molecular Pathology Core

Resource Type: core facility, service resource, access service resource

Keywords: ABRF, molecular pathology skills, confocal microscopy, image analysis, multicolor immunohistochemistry, in-situ hybridization techniques,

Availability: Restricted

Resource Name: Tulane University TNBRC Confocal Microscopy and Molecular Pathology Core Facility

Resource ID: SCR_024613

Alternate IDs: ABRF_2527

Alternate URLs: <https://coremarketplace.org/?FacilityID=2527&citation=1>

Record Creation Time: 20231024T002344+0000

Record Last Update: 20260731T043141+0000

Ratings and Alerts

No rating or validation information has been found for Tulane University TNBRC Confocal Microscopy and Molecular Pathology Core Facility.

No alerts have been found for Tulane University TNBRC Confocal Microscopy and Molecular Pathology Core Facility.

Data and Source Information

Source: [SciCrunch Registry](#)

Usage and Citation Metrics

We found 22 mentions in open access literature.

Listed below are recent publications. The full list is available at [kravitz2](#) .

Horn MD, et al. (2025) Combination antiretroviral therapy prevents SIV-induced aging in the hippocampus and neurodegeneration throughout the brain. *Journal of neurovirology*, 31(6), 485.

Kaur A, et al. (2025) Nonhuman primate model mirroring human congenital cytomegalovirus infection reveals a spectrum of vertical transmission outcomes. *Research square*.

Fredericks MN, et al. (2025) SIV/SARS-CoV-2 coinfection in rhesus macaques impacts viral shedding, host immunity, the microbiome, and viral evolution. *Frontiers in immunology*, 16, 1587688.

Wallis ZK, et al. (2025) Resolving inflammation: The impact of antiretroviral therapy on macrophage traffic in and out of the CNS. *PLoS pathogens*, 21(12), e1013180.

Xu E, et al. (2025) Spontaneous Atypical Meningioma in a Pregnant Rhesus Macaque (*Macaca mulatta*). *Journal of medical primatology*, 54(6), e70049.

Chen Y, et al. (2025) Persistent Activation of Monocytes/Macrophages and Cell Senescence in SIV-Infected Macaques on ART. *bioRxiv : the preprint server for biology*.

Reifel KM, et al. (2025) Verifying Proper Function of the Aerosol Evacuation System Prior to Sorting Potentially Infectious Samples. *Current protocols*, 5(3), e70123.

Wallis ZK, et al. (2025) Resolving Inflammation: The Impact of Antiretroviral Therapy on Macrophage Traffic In and Out of the CNS. *bioRxiv : the preprint server for biology*.

Fredericks MN, et al. (2025) SIV/SARS-CoV-2 co-infection in rhesus macaques impacts viral shedding, host immunity, the microbiome, and viral evolution. *Research square*.

Manuel TD, et al. (2025) A rhesus macaque model of congenital cytomegalovirus infection reveals a spectrum of vertical transmission outcomes. *Communications biology*, 8(1), 1647.

Van Zandt AR, et al. (2025) Simian Immunodeficiency Virus-Derived Extracellular Vesicles Induce a Chronic Inflammatory Phenotype in Healthy Astrocytes Unresolved by Anti-Retroviral Therapy. *Pharmaceutics*, 17(11).

Van Zandt AR, et al. (2025) THC reverses SIV-induced senescence in astrocytes: possible compensatory mechanism against HIV associated brain injury? *Frontiers in cellular neuroscience*, 19, 1642917.

Ellsworth CR, et al. (2024) Natural Killer Cells Do Not Attenuate a Mouse-Adapted SARS-CoV-2-Induced Disease in Rag2^{-/-} Mice. *Viruses*, 16(4).

MacLean A, et al. (2024) Combination antiretroviral therapy prevents SIV- induced aging in the hippocampus and neurodegeneration throughout the brain. *Research square*.

Currey J, et al. (2024) Upregulation of inflammatory genes and pathways links obesity to severe COVID-19. *Biochimica et biophysica acta. Molecular basis of disease*, 1870(7), 167322.

Zenere G, et al. (2024) Extracellular domain, hinge, and transmembrane determinants affecting surface CD4 expression of a novel anti-HIV chimeric antigen receptor (CAR) construct. *PloS one*, 19(8), e0293990.

Wang C, et al. (2024) Deficiency of Tlr7 and Irf7 in mice increases the severity of COVID-19 through the reduced interferon production. *Communications biology*, 7(1), 1162.

Palmer CS, et al. (2024) Non-human primate model of long-COVID identifies immune associates of hyperglycemia. *Nature communications*, 15(1), 6664.

Ellsworth CR, et al. (2024) Enhanced complement activation and MAC formation accelerates severe COVID-19. *Cellular and molecular life sciences : CMLS*, 81(1), 405.

Melton A, et al. (2024) The Impact of SIV-Induced Immunodeficiency on SARS-CoV-2 Disease, Viral Dynamics, and Antiviral Immune Response in a Nonhuman Primate Model of Coinfection. *Viruses*, 16(7).