

Resource Summary Report

Generated by T1D on Aug 5, 2026

University of Pennsylvania Perelman School of Medicine Electron Microscopy Resource Lab Core Facility

RRID:SCR_022375

Type: Tool

Proper Citation

University of Pennsylvania Perelman School of Medicine Electron Microscopy Resource Lab Core Facility (RRID:SCR_022375)

Resource Information

URL: <https://www.med.upenn.edu/electronmicroscopyresourcelab/>

Proper Citation: University of Pennsylvania Perelman School of Medicine Electron Microscopy Resource Lab Core Facility (RRID:SCR_022375)

Description: Training and service facility dedicated to providing both conventional transmission electron microscopy of cells and tissues and state of the art cryo-electron microscopy and cryo-electron tomography for structural investigation of macromolecules and cells.

Abbreviations: EMRL

Synonyms: University of Pennsylvania Perelman School of Medicine Electron Microscopy Resource Lab, Electron Microscopy Resource Lab

Resource Type: access service resource, core facility, service resource

Keywords: USEDit, ABRF, conventional transmission electron microscopy, cryo-electron microscopy, cryo-electron tomography

Availability: Open

Resource Name: University of Pennsylvania Perelman School of Medicine Electron Microscopy Resource Lab Core Facility

Resource ID: SCR_022375

Alternate IDs: ARBF_1390

Alternate URLs: <https://coremarketplace.org?citation=1&FacilityID=1390>

Record Creation Time: 20220602T050140+0000

Record Last Update: 20260805T044500+0000

Ratings and Alerts

No rating or validation information has been found for University of Pennsylvania Perelman School of Medicine Electron Microscopy Resource Lab Core Facility.

No alerts have been found for University of Pennsylvania Perelman School of Medicine Electron Microscopy Resource Lab Core Facility.

Data and Source Information

Source: [SciCrunch Registry](#)

Usage and Citation Metrics

We found 40 mentions in open access literature.

Listed below are recent publications. The full list is available at [T1D](#).

Eldaboush A, et al. (2026) Characterizing Keratinocyte-Derived Extracellular Vesicles in UVB-Irradiated Murine Skin. JID innovations : skin science from molecules to population health, 6(1), 100406.

Yoon IC, et al. (2025) Piperazine-Derived Bisphosphonate-Based Ionizable Lipid Nanoparticles Enhance mRNA Delivery to the Bone Microenvironment. Angewandte Chemie (International ed. in English), 64(3), e202415389.

Schultz K, et al. (2025) Snapshots of acyl carrier protein shuttling in human fatty acid synthase. Nature, 641(8062), 520.

Saks AJ, et al. (2025) NPF binding to Arp2 is allosterically linked to the release of ArpC5's N-terminal tail and conformational changes in Arp2/3 complex. Proceedings of the National Academy of Sciences of the United States of America, 122(8), e2421557122.

McVeigh BM, et al. (2025) Visualization of lysosomal membrane proteins by cryo electron tomography. Nature communications, 16(1), 9234.

Shimogawa M, et al. (2025) Investigation of All Disease-Relevant Lysine Acetylation Sites in ??-Synuclein Enabled by Non-canonical Amino Acid Mutagenesis. bioRxiv : the preprint server for biology.

Palumbos SD, et al. (2025) Autophagic stress activates distinct compensatory secretory pathways in neurons. Proceedings of the National Academy of Sciences of the United States of America, 122(28), e2421886122.

Chavali SS, et al. (2025) High-resolution structures of Myosin-IC reveal a unique actin-binding orientation, ADP release pathway, and power stroke trajectory. *Proceedings of the National Academy of Sciences of the United States of America*, 122(9), e2415457122.

Xu Z, et al. (2025) Cryo-electron microscopy reveals a single domain antibody with a unique binding epitope on fibroblast activation protein alpha. *RSC chemical biology*, 6(5), 780.

Han X, et al. (2025) Plug-and-play assembly of biodegradable ionizable lipids for potent mRNA delivery and gene editing in vivo. *bioRxiv : the preprint server for biology*.

Padilla MS, et al. (2025) Branched endosomal disruptor (BEND) lipids mediate delivery of mRNA and CRISPR-Cas9 ribonucleoprotein complex for hepatic gene editing and T cell engineering. *Nature communications*, 16(1), 996.

Padilla MS, et al. (2025) Solution biophysics identifies lipid nanoparticle non-sphericity, polydispersity, and dependence on internal ordering for efficacious mRNA delivery. *bioRxiv : the preprint server for biology*.

Wenger ES, et al. (2025) Structure of bifunctional variediene synthase yields unique insight on biosynthetic diterpene assembly and cyclization. *Nature communications*, 16(1), 5089.

Trigg NA, et al. (2025) Epididymal extracellular vesicles harbor and convey mRNA to sperm for transfer to zygotes. *bioRxiv : the preprint server for biology*.

Yount TA, et al. (2025) Multi-functional minor pilins coordinate type IV pilus assembly, adherence, motility, and DNA uptake in the pediatric pathogen *Kingella kingae*. *mBio*, 16(11), e0279025.

Nong J, et al. (2025) Multi-stage-mixing to control the supramolecular structure of lipid nanoparticles, thereby creating a core-then-shell arrangement that improves performance by orders of magnitude. *bioRxiv : the preprint server for biology*.

Kixmoeller K, et al. (2025) Centromeric chromatin clearings demarcate the site of kinetochore formation. *Cell*, 188(5), 1280.

Shultz RB, et al. (2025) A Nonhuman Primate Model to Evaluate Treatments for Long-Gap Ulnar Nerve Injury. *Journal of neuroscience research*, 103(8), e70069.

Liu X, et al. (2024) Evaluation of a rapid multi-attribute combinatorial high-throughput UV-Vis/DLS/SLS analytical platform for rAAV quantification and characterization. *Molecular therapy. Methods & clinical development*, 32(3), 101298.

Ren B, et al. (2024) Architecture of the *Toxoplasma gondii* apical polar ring and its role in gliding motility and invasion. *Proceedings of the National Academy of Sciences of the United States of America*, 121(46), e2416602121.