

Resource Summary Report

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CellProfiler Image Analysis Software

RRID:SCR_007358

Type: Tool

Proper Citation

CellProfiler Image Analysis Software (RRID:SCR_007358)

Resource Information

URL: <http://cellprofiler.org>

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Description: Software tool to enable biologists without training in computer vision or programming to quantitatively measure phenotypes from thousands of images automatically. It counts cells and also measures the size, shape, intensity and texture of every cell (and every labeled subcellular compartment) in every image. It was designed for high throughput screening but can perform automated image analysis for images from time-lapse movies and low-throughput experiments. CellProfiler has an increasing number of algorithms to identify and measure properties of neuronal cell types.

Synonyms: Cell Profiler, CellProfiler - cell image analysis software

Resource Type: data processing software, image analysis software, software application, software resource

Defining Citation: [PMID:21349861](#), [PMID:17076895](#), [PMID:19014601](#), [PMID:19188593](#)

Keywords: high-throughput, high content imaging, software, image, cell, phenotype, measurement, subcellular, intensity, size, shape, analysis, algorithm

Funding: NHGRI RL1 HG004671;
NIGMS R01 GM089652;
NIGMS RC2 GM092519

Availability: Free, Available for download, Freely available

Resource Name: CellProfiler Image Analysis Software

Resource ID: SCR_007358

Alternate IDs: SCR_010649, nlx_66812, nif-0000-00280

Alternate URLs: <https://sources.debian.org/src/cellprofiler/>

License: BSD License

Record Creation Time: 20220129T080241+0000

Record Last Update: 20260905T133043+0000

Ratings and Alerts

No rating or validation information has been found for CellProfiler Image Analysis Software.

No alerts have been found for CellProfiler Image Analysis Software.

Data and Source Information

Source: [SciCrunch Registry](#)

Usage and Citation Metrics

We found 3411 mentions in open access literature.

Listed below are recent publications. The full list is available at [Kravitz](#) .

Lee M, et al. (2026) LINE-1 Locus Transcription Nucleates Oncogenic Chromatin Architecture. Cancer discovery, 16(4), 800.

Lam D, et al. (2026) Effects of N361 Glycosylation on Epidermal Growth Factor Receptor Biological Function. Cancers, 18(3).

Serpieri V, et al. (2026) Bi-allelic variants in FSD1L cause a neurodevelopmental disorder overlapping with L1 syndrome. American journal of human genetics, 113(3), 600.

Huang TC, et al. (2026) Global reorganization of genome architecture at the transition to gametogenesis. Nature structural & molecular biology, 33(3), 433.

de Velasco EMF, et al. (2026) AAV-only targeting of ventral tegmental area dopamine neurons for optical self-stimulation studies in mice. bioRxiv : the preprint server for biology.

Murrey MW, et al. (2026) The myeloid SRC family kinase HCK regulates breast cancer growth by activating tumor-associated macrophage-led invasion and inhibiting cytotoxic T cell activity. Frontiers in immunology, 17, 1709102.

Zhang M, et al. (2026) Dopamine enhances surface glial glycolysis via acetylcholine and insulin/insulin-like growth factor signaling in the honeybee. iScience, 29(3), 114947.

Davis MN, et al. (2026) Parvalbumin Neuron-Targeted Loss of Alzheimer's Disease Risk Gene BIN1 Is Insufficient to Drive Cognitive or Network Excitability Changes. eNeuro, 13(3).

Yao J, et al. (2026) A multimodal study on predicting extrathyroidal extension of papillary thyroid carcinoma based on radiopathomics. *BMC endocrine disorders*, 26(1).

Mortimer AJ, et al. (2026) Comparison of AAV9-driven motor neuron transduction following different CNS-directed delivery methods in mice. *Scientific reports*, 16(1).

Sólyomvári C, et al. (2026) Time-dependent histological characterization of amyloid- β induced cholinergic and glial alterations and their modulation by dehydroepiandrosterone sulfate (DHEAS). *Frontiers in endocrinology*, 17, 1764298.

Heer CD, et al. (2026) Targeted CRISPR knockout screening identifies known and novel chemogenomic interactions between DNA damaging agents and DNA repair genes. *NAR cancer*, 8(1), zcaf052.

Wu Z, et al. (2026) Protocol for immunofluorescence detection and quantitative analysis of pH-dependent transcriptional condensates. *STAR protocols*, 7(1), 104425.

Sudakevitz-Merzbach R, et al. (2026) Temporal control of midline crossing via incoherent feedforward and feedback loops regulating Robo3. *iScience*, 29(6), 115912.

Russo MR, et al. (2026) Protocol for cell cycle-resolved high-content imaging of transcription condensate dynamics in human cells. *STAR protocols*, 7(2), 104589.

Giner-Calabuig M, et al. (2026) RecQ DNA helicases germline variants in Lynch-like syndrome. *Genetics in medicine open*, 4, 104400.

Gonzalez Torres A, et al. (2026) Spatiotemporal fluorescence imaging of microRNA activity in 3-D models of human epidermis reveals contribution of the Notch pathway in the regulation of miR-30a in aging skin. *JID innovations : skin science from molecules to population health*, 6(2), 100444.

Arbi M, et al. (2026) Mclidas localizes to centrioles and controls centriole numbers through PLK4-dependent phosphorylation. *EMBO reports*, 27(6), 1478.

Bewersdorf TN, et al. (2026) In-vitro evaluation of the proliferative and osteogenic activity of atrophic non-union derived mesenchymal stem cells compared to autologous bone graft derived mesenchymal stem cells. *European journal of trauma and emergency surgery : official publication of the European Trauma Society*, 52(1), 48.

Lasse Opsahl EL, et al. (2026) Fibroblast STAT3 Activation Drives Organ-Specific Premetastatic Niche Formation. *Cancer research*, 86(1), 22.