

Resource Summary Report

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Fred Hutchinson Cancer Center Flow Cytometry Core Facility

RRID:SCR_022613

Type: Tool

Proper Citation

Fred Hutchinson Cancer Center Flow Cytometry Core Facility (RRID:SCR_022613)

Resource Information

URL: <https://www.fredhutch.org/en/research/shared-resources/core-facilities/flow-cytometry.html>

Proper Citation: Fred Hutchinson Cancer Center Flow Cytometry Core Facility (RRID:SCR_022613)

Description: Includes services from sterile cell sorting for therapeutic applications to whole cell mass spectrometry, diagnostics and early stage clinical trials. Provides expertise from experimental design to data analysis and troubleshooting, therapeutic cell sorting. Supports isolation of cells for Phase 1 clinical trials of cellular immunotherapies.

Synonyms: Fred Hutchinson Cancer Center Flow Cytometry Shared Resource

Resource Type: service resource, access service resource, core facility

Keywords: sterile cell sorting, whole cell mass spectrometry, diagnostics, early stage clinical trials, therapeutic cell sorting, Phase 1 clinical trials, cellular immunotherapies, ABRF, USEDit, NACCSR

Availability: Open

Resource Name: Fred Hutchinson Cancer Center Flow Cytometry Core Facility

Resource ID: SCR_022613

Alternate IDs: ABRF_1533

Alternate URLs: https://coremarketplace.org/RRID:SCR_022613/?citation=1

Record Creation Time: 20220802T050144+0000

Record Last Update: 20260804T043804+0000

Ratings and Alerts

No rating or validation information has been found for Fred Hutchinson Cancer Center Flow Cytometry Core Facility.

No alerts have been found for Fred Hutchinson Cancer Center Flow Cytometry Core Facility.

Data and Source Information

Source: [SciCrunch Registry](#)

Usage and Citation Metrics

We found 50 mentions in open access literature.

Listed below are recent publications. The full list is available at [ASWG](#) .

Larsen BB, et al. (2026) Functional and antigenic constraints on the Nipah virus fusion protein. bioRxiv : the preprint server for biology.

Zhang S, et al. (2026) A CD8?? co-receptor modified to contain an intracellular CD28 signaling tail enhances TCR-engineered T cell function independent of solid-tumor-associated co-stimulatory ligands. Nature communications, 17(1), 753.

Ahn JJ, et al. (2026) Influenza hemagglutinin subtypes have different sequence constraints despite sharing extremely similar structures. bioRxiv : the preprint server for biology.

Avanessian SC, et al. (2026) IL2/IL15 Signaling Induces NK Cell Production of FLT3LG, Augmenting Anti-PD-1 Immunotherapy. Cancer immunology research, 14(1), 122.

Wan YH, et al. (2026) Monoclonal neutralizing antibodies elicited by infection with Kaposi sarcoma-associated herpesvirus reveal critical sites of vulnerability on gH/gL. PLoS pathogens, 22(1), e1013772.

Petty NE, et al. (2025) Protection of CD33-modified hematopoietic stem cell progeny from CD33-directed CAR T cells in rhesus macaques. Blood advances, 9(10), 2367.

Paatela EM, et al. (2025) A discrete region of the D4Z4 is sufficient to initiate epigenetic silencing. Human molecular genetics, 34(18), 1526.

Garcia NMG, et al. (2025) APOBEC3 Activity Promotes the Survival and Evolution of Drug-Tolerant Persister Cells during EGFR Inhibitor Resistance in Lung Cancer. Cancer research communications, 5(5), 825.

Dadonaite B, et al. (2025) Spike mutations that affect the function and antigenicity of recent KP.3.1.1-like SARS-CoV-2 variants. bioRxiv : the preprint server for biology.

Ju X, et al. (2025) Determinants of human versus mosquito cell entry by the Chikungunya virus envelope proteins. *bioRxiv : the preprint server for biology*.

Simonich CAL, et al. (2025) RSV F evolution escapes some monoclonal antibodies but does not strongly erode neutralization by human polyclonal sera. *bioRxiv : the preprint server for biology*.

Wilcox-King A, et al. (2025) Priming VRC01-precursor B cells with non-envelope immunogens disfavors boosting with HIV-1 envelope. *NPJ vaccines*, 10(1), 185.

Yu TC, et al. (2025) Pleiotropic mutational effects on function and stability constrain the antigenic evolution of influenza hemagglutinin. *bioRxiv : the preprint server for biology*.

Radford CE, et al. (2025) Comprehensive maps of escape mutations from antibodies 10-1074 and 3BNC117 for Envs from two divergent HIV strains. *Journal of virology*, 99(5), e0019525.

McCutcheon KR, et al. (2025) Recurrent Breast Cancer Cells Depend on De novo Pyrimidine Biosynthesis to Suppress Ferroptosis. *bioRxiv : the preprint server for biology*.

Radford CE, et al. (2025) Comprehensive maps of escape mutations from antibodies 10-1074 and 3BNC117 for Envs from two divergent HIV strains. *bioRxiv : the preprint server for biology*.

Simon S, et al. (2025) Design of sensitive monospecific and bispecific synthetic chimeric T cell receptors for cancer therapy. *Nature cancer*, 6(4), 647.

Simonich CAL, et al. (2025) RSV F evolution escapes some monoclonal antibodies but does not strongly erode neutralization by human polyclonal sera. *Journal of virology*, 99(7), e0053125.

Granadier D, et al. (2025) Damage-induced IL-18 stimulates thymic NK cells limiting endogenous tissue regeneration. *Nature immunology*, 26(10), 1699.

Dadonaite B, et al. (2025) Spike mutations that affect the function and antigenicity of recent KP.3.1.1-like SARS-CoV-2 variants. *Journal of virology*, 99(11), e0142325.