

Resource Summary Report

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Bio-tradis

RRID:SCR_015993

Type: Tool

Proper Citation

Bio-tradis (RRID:SCR_015993)

Resource Information

URL: <https://github.com/sanger-pathogens/Bio-Tradis>

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Description: Analysis software for the output from TraDIS (Transposon Directed Insertion Sequencing) analyses of dense transposon mutant libraries. The Bio-Tradis analysis pipeline is implemented as an extensible Perl library which can either be used as is, or as a basis for the development of more advanced analysis tools.

Abbreviations: TraDIS:Transposon Directed Insertion Sequencing

Resource Type: software toolkit, data analysis software, software application, software resource, sequence analysis software, data processing software

Defining Citation: [PMID:26794317](#), [DOI:10.1093/bioinformatics/btw022](#)

Keywords: software, tool, analysis, data, sequencing, insertion, transposon, direct, mutant, library, perl, bio.tools

Funding: Wellcome Trust WT098051;
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Availability: Free, Available for download, Freely available

Resource Name: Bio-tradis

Resource ID: SCR_015993

Alternate IDs: OMICS_11083, biotools:bio-tradis

Alternate URLs: <https://bio.tools/bio-tradis>, <https://sources.debian.org/src/bio-tradis/>

License: GNU General Public License v3.0

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Ratings and Alerts

No rating or validation information has been found for Bio-tradis.

No alerts have been found for Bio-tradis.

Data and Source Information

Source: [SciCrunch Registry](#)

Usage and Citation Metrics

We found 51 mentions in open access literature.

Listed below are recent publications. The full list is available at [ASWG](#) .

Yousief SW, et al. (2026) Genome-wide identification of tissue-specific fitness genes in murine models of Staphylococcus aureus infection. iScience, 29(1), 114261.

Di Martino ML, et al. (2025) A scalable gut epithelial organoid model reveals the genome-wide colonization landscape of a human-adapted pathogen. Nature genetics, 57(7), 1730.

Morris FC, et al. (2025) Gene dependence during mammalian Acinetobacter baumannii pneumonia and septicaemia infections. Microbial genomics, 11(11).

Harding KR, et al. (2025) Genome-wide identification of bacterial genes contributing to nucleus-forming jumbo phage infection. Nucleic acids research, 53(3).

Smallman TR, et al. (2025) Comparative analysis of shared and unique mechanisms important for diverse strains of Pasteurella multocida to cause systemic infection in mice. PLoS pathogens, 21(12), e1013398.

Yang Z, et al. (2025) Loss of cardiolipin and porins bypasses the essentiality of the sigma E cell envelope stress response in Escherichia coli. mBio, 16(9), e0161325.

Krause AL, et al. (2025) Transposon-directed insertion-site sequencing (TraDIS) analysis of Enterococcus faecium using nanopore sequencing and a WebAssembly analysis platform. Microbiology spectrum, 13(7), e0062825.

Ma Y, et al. (2025) Chromosomal genes modulating fosfomycin susceptibility in uropathogenic Escherichia coli: a genome-wide analysis. Antimicrobial agents and chemotherapy, 69(4), e0141724.

Padilla JJ, et al. (2025) Characterization of the Antibiotic and Copper Resistance of Emergent Species of Onion-Pathogenic Burkholderia Through Genome Sequence Analysis and High-Throughput Sequencing of Differentially Enriched Random Transposon

Mutants. Pathogens (Basel, Switzerland), 14(3).

Ma Y, et al. (2024) The intrinsic macrolide resistome of *Escherichia coli*. Antimicrobial agents and chemotherapy, 68(8), e0045224.

A Ghomi F, et al. (2024) High-throughput transposon mutagenesis in the family Enterobacteriaceae reveals core essential genes and rapid turnover of essentiality. mBio, 15(10), e0179824.

Alobaidallah MSA, et al. (2024) Enhancing the Efficacy of Chloramphenicol Therapy for *Escherichia coli* by Targeting the Secondary Resistome. Antibiotics (Basel, Switzerland), 13(1).

Wellner SM, et al. (2024) Genome-wide identification of fitness-genes in aminoglycoside-resistant *Escherichia coli* during antibiotic stress. Scientific reports, 14(1), 4163.

Fabian BK, et al. (2024) Identifying the suite of genes central to swimming in the biocontrol bacterium *Pseudomonas protegens* Pf-5. Microbial genomics, 10(3).

Yousief SW, et al. (2024) Optimizing phage-based mutant recovery and minimizing heat effect in the construction of transposon libraries in *Staphylococcus aureus*. Scientific reports, 14(1), 22831.

Gray J, et al. (2024) Transposon mutagenesis screen in *Klebsiella pneumoniae* identifies genetic determinants required for growth in human urine and serum. eLife, 12.

Mediati DG, et al. (2024) Genetic requirements for uropathogenic *E. coli* proliferation in the bladder cell infection cycle. mSystems, 9(10), e0038724.

Rooke JL, et al. (2024) Genome-wide fitness analysis of *Salmonella enterica* reveals *aroA* mutants are attenuated due to iron restriction in vitro. mBio, 15(10), e0331923.

Wang M, et al. (2023) Uncovering the determinants of model *Escherichia coli* strain C600 susceptibility and resistance to lytic T4-like and T7-like phage. Virus research, 325, 199048.

Ba X, et al. (2023) High-Throughput Mutagenesis Reveals a Role for Antimicrobial Resistance- and Virulence-Associated Mobile Genetic Elements in *Staphylococcus aureus* Host Adaptation. Microbiology spectrum, 11(2), e0421322.