

# Resource Summary Report

Generated by ASWG on Sep 5, 2026

## GFP-Booster\_ATTO647N

RRID:AB\_2629215

Type: Antibody

### Proper Citation

ChromoTek Cat# gba647n-100, RRID:AB\_2629215

### Antibody Information

**URL:** [http://antibodyregistry.org/AB\\_2629215](http://antibodyregistry.org/AB_2629215)

**Proper Citation:** ChromoTek Cat# gba647n-100, RRID:AB\_2629215

**Target Antigen:** eCFP, CFP, mCerulean,eGFP, wtGFP, GFP S65T, AcGFP, TagGFP, tagGFP2, sfGFP, pHluorineYFP, YFP, Venus, Citrine

**Host Organism:** alpaca

**Clonality:** monoclonal

**Comments:** Originating manufacturer of this product, tested applications: Immunofluorescence

**Antibody Name:** GFP-Booster\_ATTO647N

**Description:** This monoclonal targets eCFP, CFP, mCerulean,eGFP, wtGFP, GFP S65T, AcGFP, TagGFP, tagGFP2, sfGFP, pHluorineYFP, YFP, Venus, Citrine

**Antibody ID:** AB\_2629215

**Vendor:** ChromoTek

**Catalog Number:** gba647n-100

**Record Creation Time:** 20231110T034748+0000

**Record Last Update:** 20260830T021636+0000

### Ratings and Alerts

No rating or validation information has been found for GFP-Booster\_ATTO647N.

**Warning: Extracted Antibody Information:** "Thermo Scientific), nanobody-dye conjugate (RRID:**AB\_2629215**,"

**Extracted Specificity Statement:** "Fluorescence microscopy of the living cells is often achieved through **specific** labeling of proteins by antibody, nanobody, or bio-**specific** ligand conjugated to a fluorophore. However, most of these bio-molecules and fluorophores are not able to cross the cell membrane of a living cell, making it challenging to image intracellular proteins."

Data was mined by Antibody Watch (<https://arxiv.org/pdf/2008.01937.pdf>), from **PMID:27935478**

Originating manufacturer of this product, tested applications: Immunofluorescence

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## Data and Source Information

**Source:** [Antibody Registry](#)

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## Usage and Citation Metrics

We found 14 mentions in open access literature.

**Listed below are recent publications.** The full list is available at [ASWG](#) .

El Alaoui F, et al. (2025) Septin assemblies promote the lipid organization of membranes. Structure (London, England : 1993), 33(3), 451.

Morabito A, et al. (2025) Distinct dendritic integration strategies control dynamics of inhibition in the neocortex. Neuron.

Braun A, et al. (2023) Disynaptic inhibition shapes tuning of OFF-motion detectors in Drosophila. Current biology : CB, 33(11), 2260.

Kolabas ZI, et al. (2023) Distinct molecular profiles of skull bone marrow in health and neurological disorders. Cell, 186(17), 3706.

Bhatia HS, et al. (2022) Spatial proteomics in three-dimensional intact specimens. Cell, 185(26), 5040.

Mai H, et al. (2022) Scalable tissue labeling and clearing of intact human organs. Nature protocols, 17(10), 2188.

Park SY, et al. (2022) In vivo characterization of Drosophila golgins reveals redundancy and plasticity of vesicle capture at the Golgi apparatus. Current biology : CB, 32(21), 4549.

Klatt O, et al. (2021) Endogenous  $\gamma$ -neurexins on axons and within synapses show regulated dynamic behavior. Cell reports, 35(11), 109266.

Zhao S, et al. (2020) Cellular and Molecular Probing of Intact Human Organs. Cell, 180(4), 796.

Pan C, et al. (2019) Deep Learning Reveals Cancer Metastasis and Therapeutic Antibody Targeting in the Entire Body. *Cell*, 179(7), 1661.

Scholz N, et al. (2019) Complexin cooperates with Bruchpilot to tether synaptic vesicles to the active zone cytomatrix. *The Journal of cell biology*, 218(3), 1011.

van den Ameele J, et al. (2019) Neural stem cell temporal patterning and brain tumour growth rely on oxidative phosphorylation. *eLife*, 8.

Teng KW, et al. (2016) Labeling proteins inside living cells using external fluorophores for microscopy. *eLife*, 5.

Trovisco V, et al. (2016) bicoid mRNA localises to the *Drosophila* oocyte anterior by random Dynein-mediated transport and anchoring. *eLife*, 5.